



## The Utilization of Calcium in Watermelon Rind Extract for Osteoporosis Drug Agent

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### Abstract

Calcium is one of the essential minerals for the body, especially in the elderly and pregnant women. Using natural materials containing calcium is very important, especially natural materials that are only used as waste, such as watermelon rind. Watermelon rind has a calcium of 31 mg. Therefore, it is very efficient to be used. The manufacture of this osteoporosis drug does not refer to calcium from watermelon rind extract alone. However, it is supported by other herbal ingredients such as rhizomes (ginger, curcuma, turmeric, and fingerroot). The method of extracting calcium from watermelon rind used the destruction method. After being extracted, the extraction and some mixed with the rhizomes were tested, such as calcium levels, XRF, XRD, and FTIR tests. The result of the study showed that all samples were confirmed to have calcium. High calcium content of 13.30% was found in the watermelon rind extract sample from watermelon fruit measuring 6 kg mixed with rhizomes. All samples calcium level test results had a dominant value of 495 ppm.

**Keywords:** Calcium, osteoporosis drug, watermelon, rhizomes.

### 1. Introduction

Indonesia has a very diverse natural potential [1], known as an agricultural country, because most Indonesians work in agriculture. So, the government seeks to increase changes in agriculture that can improve people's welfare. Agriculture is changing rapidly, which could lead to a new issue with agricultural production waste that needs to be used effectively, such as fruit rind trash. One of them is the waste of fresh watermelon rind [2], which so far is still a waste that has yet to be utilized optimally [2], [3]. The primary ingredient of preference is watermelon because it is widely available and can be found throughout Indonesia, from Sabang to Merauke [4], [5]. Due to the benefits of the fruit, many farmers in Indonesia have also started growing watermelons. Fruit and seeds from the rind can be used, but they are not optimal for the rind [2].

Watermelon rind has recently shown great promise for research to enable direct application: for instance, it is employed as an antioxidant [6] in cosmetics [7]. This is where there is an update on the use of watermelon rind, which has long been utilized as an antioxidant in cosmetics. Although watermelon peel has more potassium and calcium, it also has fewer antioxidants than the fruit's flesh [5]. Calcium is the essential ingredient for use as a primary ingredient for osteoporosis drugs [6], [8]–[10]. Anybody can develop osteoporosis, but the elderly and postpartum women are particularly susceptible [8], [11].

It is known that foods like crab and lobster only have a small amount of calcium in them. However, as lobsters and crabs are pricey, it is important to look for cheaper alternatives to create calcium-containing components for osteoporosis medications. One of them is by utilizing calcium in fresh watermelon rind [2], [7]. Earlier studies showed that 100 g of watermelon rind contains 31 mg of calcium [5].

Watermelon rind extract can be stimulated by components used to make osteoporosis medications, but additional ingredients are required to maximize its support. Herbal or medicinal plants,

such as rhizomes, are employed as materials (ginger, curcuma, turmeric, and fingerroot) [1], [12]. These substances can promote the production of osteoporosis medicine ingredients since they have been proven to have molecules that are antioxidants [1], [13]–[17], antibacterial [1], [14], [16], [17], anti-inflammatory [1], [13], [14], [16], [17], and also contain some calcium [9].

## 2. Method

### 2.1. Material

The tools used for this research are a beaker, measuring cup, dropper, hot plate, funnel, spatula, petri dish, knife, blender, sample pot, oven, magnetic stirrer, digital balance, pH meter, and drawstring, regarding the ingredients used: watermelon (masses 4 kg, 5 kg, and 6 kg), rhizomes (ginger, curcuma, turmeric, and fingerroot),  $\text{HClO}_4$ ,  $\text{HNO}_3$ , solution calcium kit, distilled water, 96% alcohol, filter paper, gloves, mask, tissue, plastic wrap, laundry soap, and label paper.

### 2.2. Synthesis

There are three sections to this research. The first step is getting ready for the extraction of the watermelon rind. The watermelon rind extraction procedure using the destruction method is the second stage. The samples are ready for testing in the third stage, and some are combined with rhizome powder. [Figure 1](#) depicts a summary of the research stages.

#### 2.2.1. Preparation for Sample Extraction Process

Watermelons of all kinds are first washed, then the rind and flesh are separated, and last, the rind is shaved and dried in the hot sun until it is totally dry. Depending on the weather, the drying process takes about three days. Use a blender to mix it into a powder after drying. Along with watermelon, rhizomes, including ginger, curcuma, turmeric, and fingerroot, each weighing up to 500 g, are also available. After being washed, shaved, and dried in the sun, they are mashed in a blender to create powders of the corresponding flavors.

#### 2.2.2. Watermelon Rind Extraction Process by Destruction Method

The 45 g of watermelon rind powder was taken, placed in a beaker glass, and combined with 225 ml of  $\text{HNO}_3$ , 7.5 ml of  $\text{HCl}_4$ , and briefly agitated to ensure that the mixture was equally distributed. The beaker glass was then covered with plastic wrap and left to stand for 24 hours. The following step was heated with a hot plate at 100 °C for 10 hours after being left to stand for 24 hours. This produced a clear color precipitate (destroyed sample). Once heated, let it cool.

#### 2.2.3. Samples are Ready to be Tested and Some are Ready to be Mixed with Rhizomes

The sample from the destruction was placed in an oven at 250 °C for 1.5 hours until the brown powder was formed. The sample from the destruction was filtered using filter paper after the clear precipitate (sample) had cooled and was then washed with distilled water to pH 6 to neutralize the acid level. It is possible to evaluate or characterize brown powder using XRF, XRD, and FTIR and measure its calcium content using a calcium kit solution. Additionally, rhizomes that have been powdered are combined with 0.5 g of brown powder. A maximum of 0.5 g of each powdered rhizome was included in the combination. The next stage is to mill the brown powder and rhizome powder until they are homogeneous, after which it will be tested using the XRF, XRD, FTIR, and calcium test using a calcium kit solution.

### 2.3. Characterization

Three characterizations are used in this study's material (XRD, FTIR, and XRF) to determine the properties of each sample. The existence of calcium peaks was determined using XRD characterization. FTIR analysis to identify the functional groups that the material possesses. XRF characterization is used to determine the sample's content and its percentage composition.

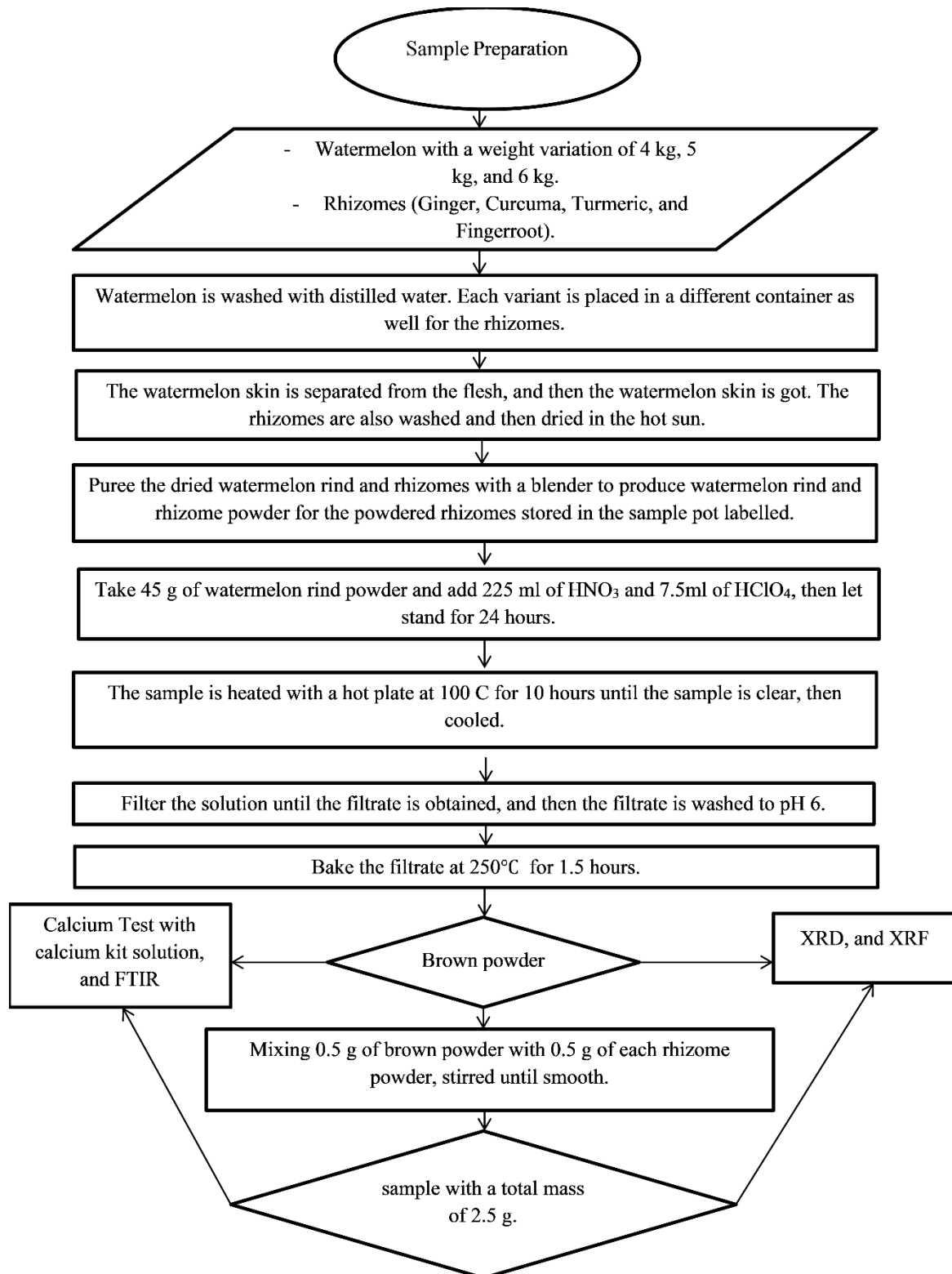
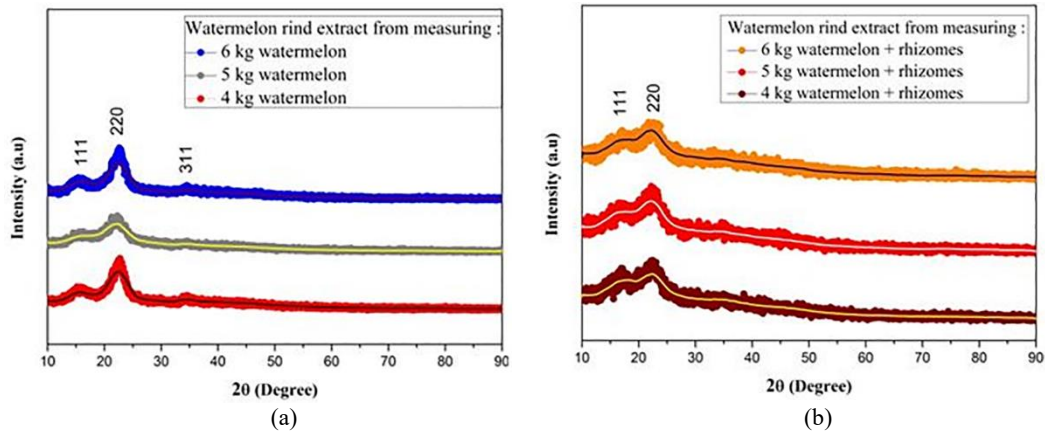


Figure 1. Research flowchart.

### 3. Result and Discussion

#### 3.1. XRD Characterization Results

The existence of calcium peaks was determined using XRD characterization. In order to smooth out the XRD data and produce a process graph, Origin software was used. Rietica software was then used to examine the plane  $hkl$ . The following are the findings from the XRD analysis of watermelon rind extract using 4 kg, 5 kg, and 6 kg watermelon variations, as shown in [Figure 2a](#).



**Figure 2.** (a) XRD patterns of watermelon rind extract from watermelon fruit masses of 4kg, 5kg, and 6 kg, (b) XRD patterns of watermelon rind extract with mixed rhizomes.

Peaks in the hkl plane are shown in [Figure 2a](#) as 111, 220, and 311. According to earlier studies, the Ca-O and Si-O functional groups are in a phase, as shown by the peaks in the 220 hkl plane [18], [19]. The Fe-O functional group is in a phase, as shown by the peak of 311 [20], [21]. According to this result, the sample that underwent XRD characterization contained Ca because its peak was in the 220 hkl planes. [Figure 2b](#) also displays the XRD results from the watermelon rind extract combined with powdered rhizomes and the XRD results from the watermelon rind extract alone (ginger, turmeric, curcuma, and fingerroot). Only the peaks in the 111 and 220 hkl fields were seen in the XRD analysis of watermelon rind extract combined with rhizomes. The findings also reveal a peak in the hkl region of 220, indicating the presence of a Ca element in the sample.

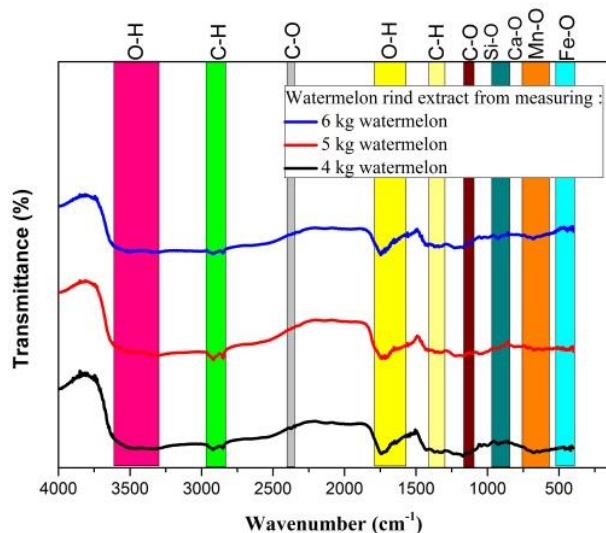
### 3.2. FTIR Results

The FTIR characterization to determine the functional groups possessed by the material. The FTIR results were analyzed by comparing the data collected with existing references. The functional groups of the watermelon rind extract shown in [Table 1](#). The FTIR results of watermelon rind extract from several variants are shown in [Figure 3](#).

Based on FTIR analysis, it is proven that the presence of the Ca functional group at wave number  $756\text{ cm}^{-1}$  is owned by the watermelon rind extract sample from watermelon fruit size 4 kg. At wave number  $765\text{ cm}^{-1}$  is owned by the watermelon rind extract sample from watermelon fruit which measuring 4 kg, there is also a wave number indicating the presence of a Ca-O functional group at a wave number of  $758\text{ cm}^{-1}$  which is owned by a sample of watermelon rind extract from a watermelon measuring 6 kg. In addition to the FTIR results from watermelon rind extract, this study also obtained FTIR results from various variants of watermelon rind extract mixed with powdered rhizomes. The presence of functional groups can be described in detail in [Table 2](#) and the spectrum of the FTIR results shown in [Figure 4](#).

**Table 1.** Functional group watermelon rind extract from watermelon fruit measuring 4 kg, 5 kg, and 6 kg.

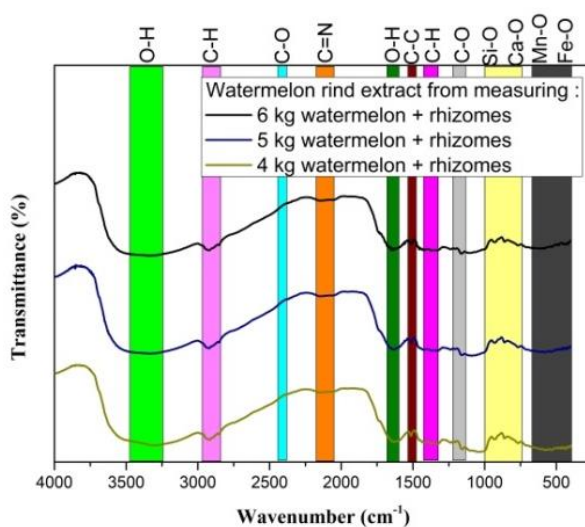
Functional groups	Wavenumbers ( $\text{cm}^{-1}$ ) in the sample of watermelon rind extract			Wavenumber ( $\text{cm}^{-1}$ ) based on references	References
	4 kg	5 kg	6kg		
Fe-O	544	550	500	400–600	[22]
Mn-O	668	673	678	500–750	[20]
Ca-O	756	765	758	750–800	[23]
Si-O	808	810	812	801–820	[23]
C-O	1092	1109	1173	1050–1200	[24]
C-H	1319	1317	1317	1300–1400	[20]
O-H	1612	1519	1700	1500–1700	[25]
C-O	2324	2322	2349	2320–2353	[26]
C-H	2850	2878	2897	2700-2900	[22]
O-H	3277	3307	3502	2930-3704	[20], [25]



**Figure 3.** FTIR spectra of watermelon rind extract from watermelon fruit masses 4 kg, 5 kg, and 6 kg.

**Table 2.** Functional group of watermelon rind extract from various variations with rhizome powder.

Functional groups	Wavenumbers (cm <sup>-1</sup> ) in the sample of watermelon rind extract mixed with rhizome powder			Wavenumber (cm <sup>-1</sup> ) based on references	References
	4 kg	5 kg	6kg		
Fe-O	532	538	530	400–600	[22]
Mn-O	607	620	611	500–750	[20]
Ca-O	785	767	765	750–800	[23]
Si-O	862	862	862	801–820	[23]
C-O	1103	1103	1161	1050–1200	[27]
C-H	1371	1368	1336	1300–1400	[20]
C-C	1514	1514	1514	1425–1550	[25]
O-H	1631	1628	1606	1500–1700	[26]
C=N	2059	2133	2056	2100–2140	[22]
C-O	2330	2324	2349	2320–2353	[20], [25]
C-H	2850	2850	2850	2700–2900	[22]
O-H	3300	3340	3342	2930–3433	[20]



**Figure 4.** Results of FTIR spectra of watermelon peel extract from various variants mixed with rhizome powder.

The results of the FTIR characterization of watermelon rind extract mixed with rhizome powder, there is also the Ca-O function group, which is found in a  $785\text{ cm}^{-1}$  wavenumber for watermelon rind extract from measuring 4 kg watermelon that is mixed with rhizomes. Furthermore, in the  $767\text{ cm}^{-1}$  wave number for a sample of watermelon rind extract from a 5 kg watermelon which was mixed with rhizome powder, and on a  $765\text{ cm}^{-1}$  wave number for a sample of watermelon rind extract from a mixed 6 kg watermelon with rhizome powder. The difference in the analysis of [Figure 3](#) and [Figure 4](#) related to the addition of rhizomes material has the effect of finding C=N and C-C groups as differentiators from the results of both FTIR spectra. From these results it shows that calcium content is true.

### 3.3. XRF Results and Calcium Level Tests

XRF characterization to find out what content is owned by the sample, and what percentage of its content. While the calcium level test is carried out to determine how large the capabilities of the calcium solubility if dissolved in solvent material such as water, testing the calcium levels indicates that the sample is feasible for medicinal ingredients that are easily dissolved if taken. The XRF test results and calcium levels of watermelon rind extract from watermelon measuring 4 kg, 5 kg, and 6 kg shown in [Table 3](#). From [Table 3](#) it is known in the sample of watermelon rind extract from a 4 kg watermelon with a calcium content of 8.6% with the solubility of 485 ppm. In watermelon rind extract from a watermelon measuring 5 kg has a calcium content of 9.1% with solubility of 490 ppm. Furthermore, in watermelon rind extract from a watermelon measuring 6 kg has a calcium content of 9.2% with solubility of 495 ppm.

In addition, this investigation generates calcium tests at watermelon rind extract mixed with rhizome powder as well as XRF data as shown in [Table 4](#) and [Table 5](#). The sample of watermelon rind extract from a 4 kg watermelon mixed with rhizome powder has a calcium content of 12.70% and a solubility of 495 ppm, and watermelon rind extract from a 5 kg watermelon mixed with rhizome powder has a calcium content of 12.90% and a solubility of 495 ppm. Furthermore, in watermelon rind extract from 6 kg watermelon mixed with rhizome powder has a calcium content of 13.30% and a solubility of 495 ppm, as shown in [Table 4](#).

**Table 3.** Results of XRF and calcium levels of watermelon rind extract from various variations.

Sample of watermelon rind extract from watermelon measuring	Elemental content (%)												Calcium level (ppm)
	Si	P	K	Ca	Sc	Ti	Cr	Mn	Fe	Ni	Cu	Yb	
4 kg	56.0	6.3	1.6	8.6	-	1.5	1.8	-	17.3	2.1	2.2	3.0	485
5 kg	66.1	6.8	1.4	9.1	-	0.9	1.1	-	10.4	1.3	2.9	-	490
6 kg	67.8	4.1	2.2	9.2	0.2	1.1	1.1	0.4	10.8	0.7	1.2	1.0	495

**Table 4.** Results of XRF characterization (Si, P, S, K, Ca, Ti, V, Cr, and Mn) and calcium levels from various variants watermelon rind extract mixed with rhizome powder.

Sample of watermelon rind extract from watermelon measuring	Elemental content (%)										Calcium level (ppm)
	Si	P	S	K	Ca	Ti	V	Cr	Mn		
4 kg	7.60	2.00	1.90	47.80	12.70	1.20	0.05	0.19	2.80	495	
5 kg	7.40	2.20	2.10	46.40	12.90	1.20	0.04	0.18	2.80	495	
6 kg	6.50	2.10	1.90	46.40	13.30	1.20	0.03	0.16	2.80	495	

**Table 5.** Results of XRF characterization (Fe, Ni, Cu, Zn, Ba, Eu, Re, Al, and Yb) and calcium levels from various variants watermelon rind extract mixed with rhizome powder.

Sample of watermelon rind extract from watermelon measuring	Elemental content (%)										Calcium level (ppm)
	Fe	Ni	Cu	Zn	Ba	Eu	Re	Al	Yb		
4 kg	20.10	0.27	1.10	0.30	0.70	0.30	0.90	-	-	495	
5 kg	19.50	0.10	0.78	0.30	0.90	0.20	1.00	1.00	0.50	495	
6 kg	21.50	0.20	0.85	0.42	0.90	0.20	0.90	-	0.50	495	

In the previous study, the calcium level of watermelon rind is 31 mg/100 g, or 0.031% [5]. While in this study, the watermelon rind extract provided the highest calcium concentration, 9.2% from 45 g, comparable to 4.14 g. In this instance, the need for more calcium is related to the requirement that osteoporosis patients ingest a maximum of 7500 mg (or 7.5 g) of calcium daily. In order to improve the calcium content, rhizome powder is used. From these results, the samples qualify to be employed as medicinal material. The maximum calcium content of watermelon rind extract combined with rhizome powder is 13.30% of 45 g, equivalent to 5.99 g. Calcium levels measured in ppm units show that calcium is easily dissolved in water, making it simpler to employ as a quickly ingested medicine.

#### 4. Conclusion

This study successfully uses the destruction method to extract calcium from watermelon rind extract, which is supported by rhizomes, a herbal substance. It has been established that the sample contains calcium based on the findings of the XRD, FTIR, XRF, and calcium testing. Watermelon rind extract from a 6 kg watermelon mixed with rhizomes has the greatest calcium level at 13.30%. The sample's calcium concentration, measured in ppm, has a dominating value of 495 ppm. According to these findings, every sample might be employed as a component in osteoporosis treatment.

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#### References

- [1] D. Kurniasari and S. Atun, "Pembuatan dan karakterisasi nanopartikel ekstrak etanol temu kunci (*boesenbergia pandurata*) pada berbagai variasi komposisi kitosan," *J. Sains Dasar*, vol. 6, no. 1, pp. 31–35, Apr. 2017, doi: [10.21831/jds.v6i1.13610](https://doi.org/10.21831/jds.v6i1.13610).
- [2] A. Aslamiah, M. Triandini, and D. R. Wicakso, "Pengambilan pektin dari albedo semangka dengan proses ekstraksi asam," *Konvers.*, vol. 3, no. 1, pp. 1–9, Apr. 2014, doi: [10.20527/k.v3i1.131](https://doi.org/10.20527/k.v3i1.131).
- [3] F. Ratnasari, Harmami, and I. Ulfin, "Efisiensi inhibisi ekstrak kulit dalam semangka sebagai inhibitor korosi tinsplate dalam media 2% NaCl," *J. Sains Seni ITS*, vol. 5, no. 1, pp. C17–C20, doi: [10.12962/j23373520.v5i1.15586](https://doi.org/10.12962/j23373520.v5i1.15586).
- [4] I. K. Adnyana, N. D. Arlinda, and D. Safitri, "Efek antilelah ekstrak air mesokarp semangka kuning (*Citrullus lanatus* Thunb.) tanpa biji," *Kartika: J. Ilm. Farm.*, vol. 2, no. 2, pp. 1–6, Dec. 2014, doi: [10.26874/kjif.v2i2.27](https://doi.org/10.26874/kjif.v2i2.27).
- [5] F. Andrianto, "Pengaruh sari kulit dan buah semangka merah (*Citrullus lanatus*) sebagai bahan pengencer terhadap motilitas dan viabilitas spermatozoa domba," B.S. thesis, Dept. Anim. Health, Airlangga University, Surabaya, Indonesia, 2016.
- [6] G. Gladvin, G. Sudhaakr, V. Swathi, and K. V. Santhisri, "Mineral and vitamin compositions contents in watermelon peel (rind)," *Int. J. Curr. Microbiol. Appl. Sci.*, vol. 5, no. 5, pp. 129–133, Oct. 2017.
- [7] N. L. P. S. Ekayanti, F. L. Darsono, and S. Wijaya, "Formulasi sediaan krim pelembab ekstrak air buah semangka (*Citrullus lanatus*)," *J. Farm. Sains Terap.*, vol. 6, no. 1, pp. 38–45, Feb. 2019, doi: [10.33508/jfst.v6i1.2011](https://doi.org/10.33508/jfst.v6i1.2011).
- [8] E. Kristiningrum, "Farmakoterapi untuk osteoporosis," *Cermin Dunia Kedokt.*, vol. 47, no. 5, pp. 41–48, 2020.
- [9] S. Nadia and A. S. Daulay, "Kandungan kalsium rimpang kunyit sebagai makromineral yang bermanfaat bagi kesehatan," in *Pros. Sem. Nas. Expo II Has. Penelit. Pengabd. Masy. 2019*, vol. 2, no. 2, pp. 855–859, Sep. 2019.
- [10] D. Rahmelia, A. W. M. Diah, and I. Said, "Analisis kadar kalium (K) dan kalsium (Ca) dalam kulit dan daging buah terung kopek ungu (*Solanum melongena*) asal Desa Nupa Bomba Kecamatan Tanantovea Kabupaten Donggala," *J. Akademika Kim.*, vol. 4, no. 3, pp. 143–148, Aug. 2015.
- [11] R. E. Brenneman, "Osteoporosis, the silent disease: Prevention and treatment of fragility fractures in a structured program" *J. Lanc. Gen. Hosp.*, vol. 11, no. 4, pp. 112–116, 2016.

- [12] H. Mulyani, S. H. Widyastuti, and V. I. Ekowati, "Tumbuhan herbal sebagai jamu pengobatan tradisional terhadap penyakit dalam serat primbon jampi jawi jilid I," *J. Penelit. Hum.*, vol. 21, no. 2, pp. 73–91, Okt. 2016, doi: [10.21831/hum.v21i2.13109](https://doi.org/10.21831/hum.v21i2.13109).
- [13] I. K. Adnyana, "Kajian efektivitas penggunaan tanaman obat dalam jamu untuk pengobatan osteoporosis," *J. Farm. Galenika*, vol. 3, no. 1, pp. 20–29, Mar. 2017.
- [14] S. Handayani, S. Mursiti, and N. Wijayati, "Uji aktivitas antibakteri senyawa flavonoid dari rimpang temu kunci (*Kaempferia pandurata* Roxb.) terhadap streptococcus mutans," *Indones. J. Chem. Sci.*, vol. 7, no. 2, pp. 146–152, Sep. 2018, doi: [10.15294/IJCS.V7I2.20810](https://doi.org/10.15294/IJCS.V7I2.20810).
- [15] T. Irianti *et al.*, "Pembuatan sediaan tabir surya ekstrak etanol rimpang temu kunci (*Boesenbergia pandurata* (Roxb.) Schlecht), aktivitas inhibisi fotodegradasi tirosin dan kandungan fenolik totalnya," *Maj. Farm.*, vol. 16, no. 2, pp. 218–232, Jun. 2020, doi: [10.22146/farmaseutik.v16i2.49421](https://doi.org/10.22146/farmaseutik.v16i2.49421).
- [16] M. Rahminiwati, S. Rahmatullah, I. Batubara, and S. S. Achmadi, "Potency of turmeric rhizome extract as prebiotic agent for *Lactobacillus plantarum* growth promoter in vitro," *J. Ilm. Kefarm. Indones.*, vol. 12, no. 1, pp. 37–42, Apr. 2014.
- [17] C. Y. Shan and Y. Iskandar, "Studi kandungan kimia dan aktivitas farmakologi tanaman kunyit (*Curcuma longa* L.)," *Farmaka*, vol. 16, no. 2, pp. 547–555, Aug. 2018, doi: [10.24198/jf.v16i2.17610](https://doi.org/10.24198/jf.v16i2.17610).
- [18] E. Hernawan, "Kandungan mineral makro (natrium dan kalsium) dalam air zamzam yang beredar di kota Tasikmalaya," *J. Kesehat. Bakti Tunas Husada: J. Ilm. Ilm. Keperawat., Analis Kesehat. Farm.*, vol. 18, no. 1, pp. 98–104, Feb. 2018, doi: [10.36465/jkbth.v18i1.310](https://doi.org/10.36465/jkbth.v18i1.310).
- [19] K. Adebowale, A. Egbedina, and B. Shonde, "Adsorption of lead ions on magnetically separable Fe<sub>3</sub>O<sub>4</sub> watermelon composite," *Appl. Water Sci.*, vol. 10, pp. 1–8, Sep. 2020, doi: [10.1007/s13201-020-01307-y](https://doi.org/10.1007/s13201-020-01307-y).
- [20] N. Mufti *et al.*, "The effect of growth temperature on the characteristics of ZnO Nanorods and its optical properties," *J. Phys.: Conf. Ser.*, vol. 1057, no. 1, p. 012005, Jul. 2018, doi: [10.1088/1742-6596/1057/1/012005](https://doi.org/10.1088/1742-6596/1057/1/012005).
- [21] A. S. Rini *et al.*, "Structural and morphological studies of silver nanoparticles prepared using *Citrullus lanatus* rind extract," *AIP Conf. Proc.*, vol. 2320, no. 1, p. 030010, Mar. 2021, doi: [10.1063/5.0037960](https://doi.org/10.1063/5.0037960).
- [22] S. Bahtiar *et al.*, "Synthesis, investigation on structural and magnetic behaviors of spinel M-ferrite [M= Fe; Zn; Mn] nanoparticles from iron sand," *IOP Conf. Ser.: Mater. Sci. Eng.*, vol. 202, no. 1, p. 012052, May 2017, doi: [10.1088/1757-899X/202/1/012052](https://doi.org/10.1088/1757-899X/202/1/012052).
- [23] B. Budiman and A. Dwi, "Sintesis keramik kalsium silikat menggunakan kalsium karbonat (CaCO<sub>3</sub>) dan silikon dioksida (SiO<sub>2</sub>) dengan teknik reaksi padatan pada suhu sintering 1200° C," *J. Teori Aplikasi Fis.*, vol. 1, no. 1, pp. 1–5, Jan. 2013, doi: [10.23960%2Fjtaf.v1i1.466](https://doi.org/10.23960%2Fjtaf.v1i1.466).
- [24] K. S. Bejigo, B. J. Park, J. H. Kim, and H. H. Yoon, "Synthesis and evaluation of graphene aerogel-supported Mn<sub>x</sub>Fe<sub>3-x</sub>O<sub>4</sub> for oxygen reduction in urea/O<sub>2</sub> fuel cells," *ChemistryOpen*, vol. 8, no. 5, pp. 615–620, May 2019, doi: [10.1002/open.201900105](https://doi.org/10.1002/open.201900105).
- [25] S. T. Yazdi, P. Iranmanesh, S. Saeednia, and M. Mehran, "Structural, optical and magnetic properties of Mn<sub>x</sub>Fe<sub>3-x</sub>O<sub>4</sub> nanoferrites synthesized by a simple capping agent-free coprecipitation route," *Mater. Sci. Eng.: B*, vol. 245, pp. 55–62, Jun. 2019, doi: [10.1016/j.mseb.2019.05.009](https://doi.org/10.1016/j.mseb.2019.05.009).
- [26] S. Kristianingrum. *Handout Spektroskopi Infra Merah (Infrared Spectroscopy, IR)* [Online]. Yogyakarta, Indonesia: Universitas Negeri Yogyakarta, 2016. Available: <http://staff.uny.ac.id/sites/default/files/pendidikan/Susila%20Kristianingrum,%20Dra.,%20M.Si/Handout-INSTRUMEN-IR-Susi.pdf>.
- [27] E. Monica and R. Rollando, "Identifikasi dan isolasi senyawa likopen dari semangka (*Citrullus Lanatus*)," *J. Ilm. Farm. Farm. Klin.*, vol. 16, no. 1, pp. 80–85, Jun. 2019, doi: [10.31942/jiffk.v16i01.2933](https://doi.org/10.31942/jiffk.v16i01.2933).